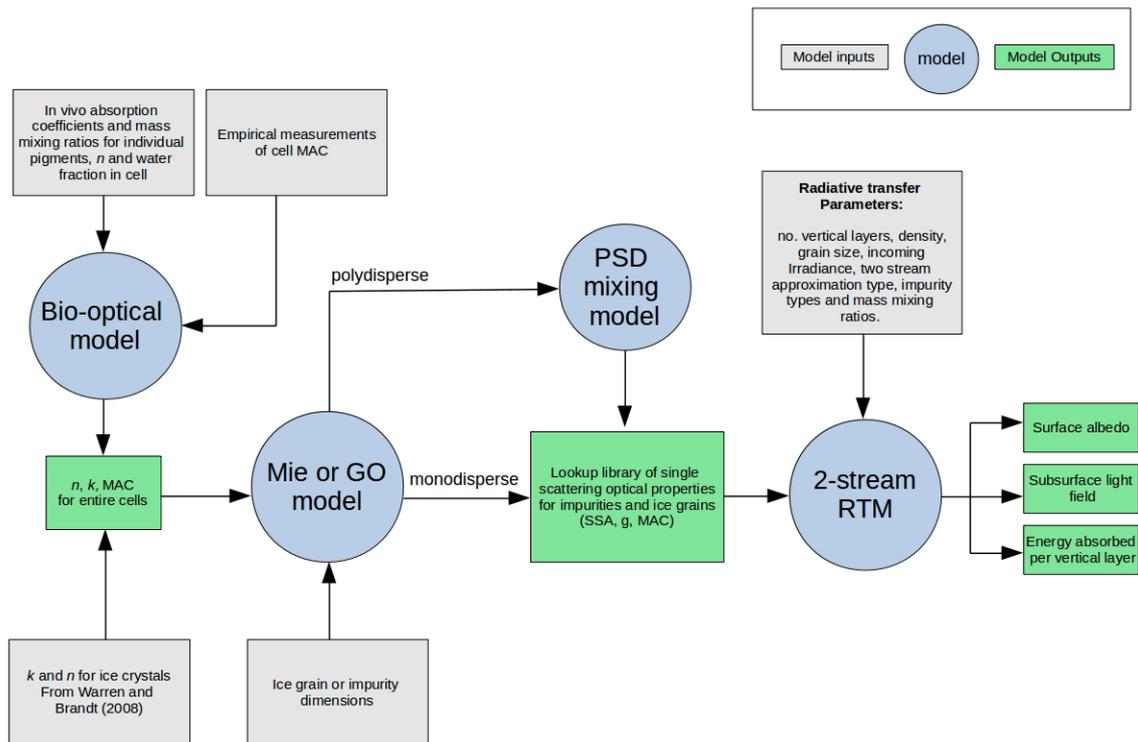


The Cryosphere

**Cook et al. (2019): Glacier Algae accelerate melt rates on the southwestern Greenland Ice Sheet**

Revised Supplementary Information

# S1: Schematic diagram of the BioSNICAR\_GO model structure

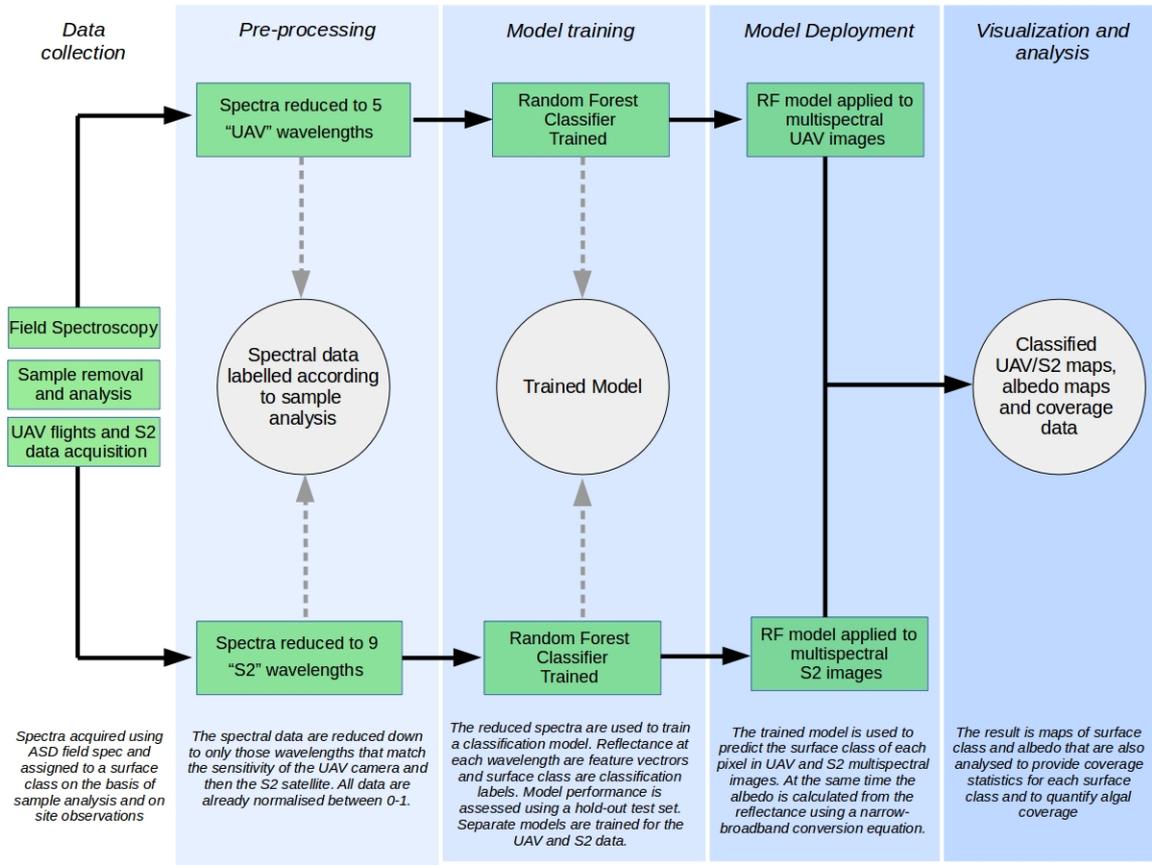


## **Supp Info 2: Mineral dust sampling and particle size distribution (PSD).**

High algal biomass ice samples were collected in sterile sample bags and melted at ambient temperatures (5-10 °C). The thawed samples were filtered onto glass fiber filters (0.7 μm pore size), from which the solids were removed into a glass jar using a stainless steel spatula. In 50 mL centrifuge tubes, the samples were treated using 30% H<sub>2</sub>O<sub>2</sub> (w/w) (Honeywell Fluka™) to remove the organic fraction. The samples (1-2 g) were sonicated (VWR ultrasonic cleaner) in 45 mL of the H<sub>2</sub>O<sub>2</sub> treatment for 10 min to disaggregate the material. The samples were left in the H<sub>2</sub>O<sub>2</sub> treatment for 48 h, after which they were centrifuged for 10 min at 4000 rpm (Eppendorf centrifuge 5810). The supernatant was removed, and the H<sub>2</sub>O<sub>2</sub> solution was replaced. This process was repeated up to ten times until no more organic oxidation was observed. The remaining mineral fraction was washed three times in water (Sartorius arium<sup>®</sup> pro ultrapure water), with centrifugation after each wash.

A 5 mg of H<sub>2</sub>O<sub>2</sub>-treated sample was suspended in 10 mL of ultrapure water. The sample was sonicated to disaggregate the grains. The suspension was dispersed onto a 0.2 μm polycarbonate filter (Sartorius Track-Etch Membrane, 0.2 μm). Once dry, a section of each filter was adhered to a stainless steel SEM stub using an adhesive carbon tab. The sample was coated with 8 nm of Ir (Agar high resolution sputter coater). The PSD was determined using a Zeiss Ultra Plus field emission scanning electron microscope (FE-SEM) operated at 20 kV. Automated particle counting software was used to determine the PSD in an area of approximately 1 mm<sup>2</sup>.

### Supp Info 3: Schematic diagram of the classification method



**Supp Info 4: A) Performance metrics for supervised classification algorithms on training data using five bands coincident with MicaSense Red-Edge multispectral imagery, plus the final model performance on the test set; B) Performance metrics for supervised classification algorithms on training data using eight bands coincident with Sentinel-2 multispectral imagery, plus the final model performance on the test set.**

**A:**

<b>Model</b>	<b>Accuracy</b>	<b>Precision</b>	<b>Recall</b>	<b>F1 Score</b>
K-Nearest Neighbours	0.90	0.74	0.78	0.76
Naive-bayes	0.90	0.80	0.81	0.80
Support Vector Machine	0.94	0.89	0.87	0.88
Random Forest	0.99	0.99	0.95	0.97
Ensemble	0.92	0.76	0.81	0.78
<b>RF performance on test set</b>	<b>0.90</b>	<b>0.91</b>	<b>0.90</b>	<b>0.90</b>

**B:**

<b>Model</b>	<b>Accuracy</b>	<b>Precision</b>	<b>Recall</b>	<b>F1 Score</b>
K-Nearest Neighbours	0.89	0.90	0.89	0.87
Naive-bayes	0.89	0.89	0.89	0.89
Support Vector Machine	0.96	0.96	0.96	0.96
Random Forest	0.99	0.99	0.99	0.99
Ensemble	0.93	0.93	0.93	0.93
<b>RF performance on test set</b>	<b>0.92</b>	<b>0.93</b>	<b>0.92</b>	<b>0.93</b>

**Supp Info 5: A) Hourly radiative forcing for  $H_{\text{bio}}$  and  $L_{\text{bio}}$  ice; B) Mea depth of absorption feature for  $H_{\text{bio}}$ ,  $L_{\text{bio}}$ , CI and SN sites; C) p-values for spectral Bonferroni-corrected t-tests for albedo between each surface class; D) t-statistics for spectral Bonferroni-corrected t-tests for albedo between each surface class.**

