

Interactive comment on “Effects of the interaction of ocean acidification, solar radiation, and warming on biogenic dimethylated sulfur compounds cycling in the Changjiang River Estuary” by Shan Jian et al.

Shan Jian et al.

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Received and published: 4 February 2018

Dear Editor and Reviewers, Thank you for your useful comments and suggestions on the language and structure of our manuscript (bg-2017-453). The manuscript has been carefully revised according to reviewers' comments and polished by the polishing company. The following are the reviewer's comments related to the manuscript and how we have addressed each of reviewer's concerns (red words). Changes have been marked as red in the manuscript.

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Question 1. On the core. 1.1 Questionable methodological approach The methodological approach used by the authors is incomplete and raises several concerns. The most alarming one is trying to report the impacts of OA on the dynamics of biogenic sulphur compounds and having only this to say about carbonate system measurements and monitoring throughout the 23-day experiment: p.3 Lines 29-30 “Seawater pH was constantly measured using a pH meter (. . .) and the precision is +/- 0.002.” Have the authors taken into account a correction factor based on fluctuating temperature? pH meters are usually calibrated at 25°C and substantial variation in measurements can ensue from variability in temperature. The authors mention that “. . .temperature was continuously controlled by circulating in situ seawater, hot water, and ice.”, suggesting that potentially significant variation in temperature occurred during 23 days. Have they monitored salinity throughout the experiment? The pH scale (National Bureau of Standard scale (pHNBS), free scale (pHF), total scale (pHT), or seawater scale (pHSWS)) is not mentioned in the manuscript.

Reply: Thanks for the reviewer’s comment. Yes, we have taken into account a correction factor based on fluctuating temperature. pH meters were calibrated at 25 °C in the determination and three parallel samples were measured for each pH value. Temperature was continuously controlled by circulating in situ seawater, hot water, and ice in the temperature experiments for 8 h (not the incubation experiment of ocean acidification for 23 days). In the incubation experiment of ocean acidification (23 days), all incubation barrels were exposed to sunlight in a flow-through water bath to maintain treatments at their in situ temperature. Unfortunately, the salinity data were not determined throughout the experiment. According to the reviewer’s suggestion, we have added the relevant content in the revised manuscript. “Seawater pH was continually measured using a pH meter (S210 SevenCompact™, Mettler, Germany, precision: ± 0.002), using the National Bureau of Standards (NBS) scale to calibrate the pH electrode.” (Page 4 Line 9-11)

Question: To ensure reproducibility, it is critical to report and monitor at least two vari-

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ables of the carbonate system of seawater (as well as salinity and temperature) for the entire period of the experiment. The authors do add a separate paragraph at the end of the Analytical procedures section where they mention the following: p.5 Lines 9-12 “Total dissolved inorganic carbon (DIC) was determined by a DIC analyzer (AS-C2, Apollo SciTech Inc., Georgia, USA). 10 A sample of 0.5 mL was acidified by 0.5 mL 10

Reply: Thanks for the reviewer’s comment. Unfortunately, no temporal monitoring is available in our experiment. DIC was measured only at the beginning of the incubation experiment and the Standard Deviations have been added in Table 1 in (Page 31 Line 1).

A volume of 0.5 mL is sufficient for the measurement of DIC. We have added references in the revised manuscript. “Cai, W. J., Dai, M., Wang, Y., Zhai, W., Huang, T., Chen, S., Zhang, F., Chen, Z., Wang, Z.: The biogeochemistry of inorganic carbon and nutrients in the Pearl River estuary and the adjacent Northern South China Sea. *Contin. Shelf Res.*, 24, 1301–1319, <https://doi.org/10.1016/j.csr.2004.04.005>, 2004” (Page 15 Line 25-27) “Cai, W. J., Wang, Y.: The chemistry, fluxes, and sources of carbon dioxide in the estuarine waters of the Satilla and Altamaha Rivers, Georgia, *Limnol. Oceanogr.*, 43, 657–668, <https://doi.org/10.4319/lo.1998.43.4.0657>, 1998.” (Page 15 Line 28-29)

Question: Furthermore, the authors do not mention the formulations used to calculate all variables with CO2SYS: concentrations of total boron, CO2 solubility (K_0), Dissociation constants of carbonic acid (K_1 and K_2), boric acid (K_b), water (K_w), phosphoric acid (K_{p1} , K_{p2} , K_{p3}), silicic acid (K_{si}), hydrogen fluoride (K_f), and bisulfate (K_s), Solubility products of calcite (K_{spc}) and aragonite (K_{spa}). A very useful document prepared in the framework of the data management activity of the Ocean Acidification International Coordination Centre of the International Atomic Energy Agency can be found here (OAICCL: www.iaea.org/ocean-acidification) and shares recommendations proposed in the Guidelines for reporting ocean acidification data in scientific journals. I also worry about trying to measure concentrations of volatile DMS within non collapsible barrels from which volumes of water are extracted daily creating more and more

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headspace over the course of 23 days (figure 3). The authors do not offer any explanation for this, or any suggestions as to how they correct for this problem. We learn that the authors used filtered (what porosity? Axenic? 0.2 μ m, 0.7 μ m?) water to run side-incubations to measure the impact of light on DMS. A first thought here is that any type of filtration process may introduce biases. DMSP-producing communities are notoriously sensitive to filtration, the most common problem is the pressure-induced heightening of the DMSPd pool caused by the rupture of healthy cells. This can artificially enhance pools of DMS through mixing of DMSPd and DMSP-lyase enzymes. This is never mentioned. Also, DMS is controlled by both biology (bacteria, viruses, phyto, zoo) and physics (light, wind). Thus the response of DMS to various light regimes may occur through various pathways (photochemically but also through its impact on primary producers or bacterioplankton). The methodological scheme suggests that the authors here are only investigating the impact of pH and light on photolysis rates of DMS. Yet it is unclear if bacteria/phyto are still present in these experiments (what is the porosity of the filter used?). On page 8 line 25 the authors state the following: "In order to assess the community-level response to the ocean change in future CO₂ and light conditions, photolysis rate constants (K , d^{-1}) for DMS were calculated." How is it possible to assess a "community-level" response when filtered water is used presumably to focus on physical processes only (photochemistry)? The authors state objectives that cannot be answered through the methods they use.

Reply: Thanks for the reviewer's comment. We agree with the reviewer's comment. CO₂SY: The value of K_0 (the solubility coefficient of CO₂) and the conversion between the fugacity and the partial pressure of CO₂ are from Weiss, R. F., *Marine Chemistry* 2:203-215, 1974. The value of K_b (for boric acid), in constant choices 1 to 5, is from Dickson, Andrew G., *Deep-Sea Research* 37:755-766, 1990. The boron concentration in constant choices 1 to 5 is from Uppstrom, Leif, *Deep-Sea Research* 21:161-162, 1974. Values of K_W (for H₂O), K_{P1} , K_{P2} , and K_{P3} (for phosphoric acid), and K_{Si} (for silicic acid) are from (in constant choices 1 to 5) Millero, Frank J., *Geochemica et Cosmochemica Acta* 59:661-677, 1995. K_1 and K_2 of carbonic acid are from the data

that refit by Dickson and Millero (1987) on the seawater scale. In the present study, we measured the DMS concentration in solution after the gas-liquid equilibrium. Changes in DMS production by phytoplankton will result in a simultaneous change in DMS concentration in the solution after the equilibration between solution and headspace, so the presence of the headspace will not affect the overall trend of DMS concentration changes. The seawater using in the photodegradation experiment was filtered with 0.4 and 0.2 μm Whatman GF/F filter and all equipment were sterilized during the experiment. In addition, the DMS samples were collected and measured without any filtration process. We have added the relevant content in the revised manuscript. “In the separate photodegradation experiment, 1 L seawater sample was filtered directly into precleaned Qorpak bottles through 0.45 and 0.2 μm polyethersulfone membranes to remove bacteria, phytoplankton, zooplankton and original DMS. Then the standard solution of DMS was added to the filtered seawater.” (Page 4 Line 12-14) Furthermore, we have rephrased the sentence “In order to assess the community-level response to the ocean change in future CO₂ and light conditions, photolysis rate constants (K , $d-1$) for DMS were calculated” in the revised manuscript. “In order to assess the photodegradation process of DMS response to the ocean change in future CO₂ and light conditions, photolysis rate constants (K , $d-1$) for DMS were calculated.” (Page 9 Line 18-19)

Question: The authors use 20L barrels to run their 23 day incubation experiment. Of those 23 days we learn that 18 are onboard the ship and 5 are conducted in the lab (page 3 line 16)? This is very worrisome. No further information is given here. How were the barrels moved? Presumably a lot of mixing occurred during the transit? How were temperatures kept constant during this time? For an experiment looking to identify the impacts of light and temperature, (and pH) on the dynamics of a DMSP-DMS-producing community, this major shift in environmental settings introduces a lot of potential variability that can obscure the response. Yet this is simply glanced over and never mentioned again. Also, I am curious to know (this could be calculated from the methodologies) how much water was left in the 20L barrels prior to the last sampling

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day? At least 10L? Or less?

Reply: The barrels were moved by a truck. The barrels were covered with lids throughout the experiment, including the transportation process, which can avoid the gas loss caused by mixing. The relevant content has been added in the manuscript. “All barrels were sealed with lids throughout the experiment, including the transportation process”. (Page 4 Line 6-7) The transport took about twelve hours, during which time the temperature was controlled by the seawater in the incubation tank to keep the incubation barrels at the same temperature. By calculating, approximately 11.6 L seawater was left in the 20 L barrels on the last sampling day.

Question: 1.2 The authors treat the response of a complex Estuary to OA the same way they would the response of open waters to OA. The OA problem is increasingly complex in estuarine and coastal waters where freshwater runoff, tidal mixing and high biological activity contribute to variations in CO₂ and pH on different time scales. The surface mixed-layer pCO₂ can vary spatially and is strongly modulated by biological productivity during the phytoplankton growth season. Surface pHT in Estuaries can also vary significantly within a single tidal cycle, nearly as much as the world’s ocean have experienced in response to anthropogenic CO₂ uptake over the last century. .
. Studying the impact of OA in these circumstances (high natural variability in pH and possible resilience of communities) is not devoid of interest, but the authors do not state this, they don’t even mention these natural fluctuations and how these could affect the communities, on a basic level, are these communities already tolerant to rapid fluctuations in pH? They treat the impact of OA on this very complex ecosystem as if it was the impact of OA on open oceanic waters. This is troublesome.

Reply: Thanks for the reviewer’s comment. We agree with the reviewer’s comment and have added relevant content in the revised manuscript. “In recent years, the effects of simulated future CO₂-induced seawater acidification on DMS production are still controversial (Avgoustidi et al., 2012; Kerrison et al., 2012; Kim et al., 2010; Vogt et al., 2008) and very few data are available on the OA effects on DMS production in

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estuaries. The patterns of low pH states observed in complex estuary are largely due to the result of freshwater runoff, natural mixing and high biological activity. By the end of this century, OA may become the dominant process reducing the pH in estuary area (Feely et al., 2010). Studying the impact of OA on the DMS cycle in estuary is very necessary and meaningful (Reum et al., 2015).” (Page 3 Line 15-20). “Feely, R. A., Alin, S. R., Newton, J., Sabine, C. L., Warner, M., Devol, A., Krembs, C., Maloy, C.: The combined effects of ocean acidification, mixing, and respiration on pH and carbonate saturation in an urbanized estuary. *Estuar. Coast. Shelf Sci.*, 88, 442–449, <https://doi.org/10.1016/j.ecss.2010.05.004>, 2010.” (Page 16 Line 21-23) In the acidification incubation experiment, chrysophyceae and cryptophyceae disappeared during HC treatment at the late stage of the experiment, indicating a great influence of OA on their growth. But they made up a smaller percentage of this community and diatoms and dinoflagellates dominated the phytoplankton community. According the result of section 4.1.3, the ratios of DMS/Chl-a, DMSPt/Chl-a, and DMSOt/Chl-a showed no difference in the dimethylated sulfur compounds production in terms of phytoplankton biomass. This result indicated that the relative strength of DMS, DMSP, and DMSO production by the phytoplankton communities has not changed and these communities were tolerant to rapid fluctuations in pH. In order to better understand, we added relevant content in the revised manuscript. “This result indicated that the productions of dimethylated sulfur compounds per unit biomass were not affected by pH perturbations and these communities were tolerant to rapid fluctuations in pH.” (Page 12 Line 29-31)

Question: 1.3 Statistical approach The description of the statistical strategies used by the authors is confusing and their application to the dataset is questionable. The title of the paper is full of promise “Effects of the interaction of OA, solar radiation and warming on biogenic sulphur compounds cycling. . .” but fails to deliver on that promise. The methodological and statistical approach proposed by the authors does not allow them to explore potential interactive effects of three stressors on the sources and sinks of DMS and DMSP. Multiple stressors can influence a variable independently (additive), or interact to either reduce (antagonistic) or enhance (synergistic) that variable in a non-

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linear, unpredictable fashion. I suggest the authors read Todgham and Stillman (2013), Riebesell and Gattuso (2015), Reum et al. (2015), Gunderson et al (2016). A collapsed factorial approach would have been more informative and simpler to interpret, see Boyd et al (2015). Please also see: “SCOR working group 149 <https://scor149-ocean.com> Changing Ocean Biological Systems (COBS): How will biota respond to a changing ocean?” It contains pertinent information for moving studies from single to multiple drivers.

Reply: In this study, we explored the potential interactive effects of ocean acidification and solar radiation or warming, the title of the paper may be improperly expressed. Thanks for the reviewer’s comment and we have changed the title and related narration in the revised manuscript. “Effects of the interaction of ocean acidification and solar radiation/warming on biogenic dimethylated sulfur compounds cycling in the Changjiang River Estuary” (Page 1 Line 1-3) “Moreover, we examined the photolysis rate and concentration changes of DMS under the dual stressors of changing pH and solar radiation/temperature and assessed the coupling effects of OA and solar radiation/warming on biogenic dimethylated sulfur compounds cycling.” (Page 3 Line 22-23) “3.2 DMS and DMSP concentration response to the interaction among OA and solar radiation/warming” (Page 9 Line 15) Thanks to the references recommended by the reviewer. We have added these references and relevant interpretation in the revised manuscript. “So these concomitant global change variables should be taken into consideration to better explain the effect of acidification on DMS (Todgham and Stillman, 2013; Boyd et al., 2015; Riebesell and Gattuso, 2015; Gunderson et al., 2016).” (Page 3 Line 14) “Studying the impact of OA on the DMS cycle in estuary is very necessary and meaningful (Reum et al., 2015)” (Page 3 Line 19-20) “Todgham, A. E., Stillman, J. H.: Physiological responses to shifts in multiple environmental stressors: relevance in a changing world, *Integr. Comp. Biol.*, 53, 539–544, <https://doi.org/10.1093/icb/ict086>, 2013.” (Page 20 Line 9-10) “Boyd P W, Lennartz S T, Glover D M, Doney, S. C.: Biological ramifications of climate-change-mediated oceanic multi-stressors, *Nat. Clim. Change*, 5, 71–79, <https://doi.org/10.1038/nclimate2441>, 2015.” (Page 15 Line 18-

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19) “Riebesell, U., Gattuso, J. P.: Lessons learned from ocean acidification research, *Nat. Clim. Change*, 5, 12–14, <https://doi.org/10.1038/nclimate2456>, 2015.” (Page 19 Line 13-14) “Gunderson, A. R., Armstrong, E. J., Stillman, J. H.: Multiple stressors in a changing world: the need for an improved perspective on physiological responses to the dynamic marine environment, *Ann. Rev. Mar. Sci.*, 8, 357–378, <https://doi.org/10.1146/annurev-marine-122414-033953>, 2016.” (Page 17 Line 6-8) “Reum J C P, Alin S R, Harvey C J, Bednaršek, N., Evans, W., Feely, R. A., Hales, B., Lucey, N., Mathis, J. T., McElhany, P., Newton, J. Sabine, C. L.: Interpretation and design of ocean acidification experiments in upwelling systems in the context of carbonate chemistry co-variation with temperature and oxygen. *ICES J Mar. Sci.*, 73, 582–595, <https://doi.org/10.1093/icesjms/fsu231>, 2015.” (Page 19 Line 9-12) “In addition, Todgham and Stillman (2013) reported that multiple stressors can influence performance independently (additive) or interact interactively (antagonistic or synergistic). We can infer that OA and warming play a synergistic role in the production of dimethylated sulfur compounds. This result is consistent with that reported by Six et al. (2013) who showed that global warming can be amplified through pH dependence of DMS production.” (Page 13 Line 21-25)

Question: On page 12 lines 24-26 the authors mention the following: “The effect of the interaction between OA and environmental conditions complicates the overall ecosystem response. Hence, comprehensive consideration of OA and solar radiation can better interpret and understand feedbacks between OA and global climatic change.” A very small exploration of interactive effects is shown in Figure 6 for pH and temperature, and there appears to be a subset of information on pH and certain measurements of light in Table 3, but the interactive impacts of solar radiation, pH and temperature together are not explored. I do not see any “comprehensive consideration”. Table 3 seems to relate one type of pH with one type of light treatment, but no combinations. . . pH 8.1 + light control (quartz), pH 8.1 + Mylar D (UVA), pH8.1 + Plexiglass (PAR), pH8.1 + Dark. pH + light control (quartz), pH 8.1 + Mylar D (UVA), pH8.1 + Plexiglass (PAR), pH8.1 + Dark. Same with pH 7.9 or 7.7 with various light treatments. It is not

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clear what the rationale is behind choosing a specific pH with a specific light treatment (?).

Reply: According to the reviewer's suggestion, we have reworded this sentence in the revised manuscript. "Hence, consideration of the interaction between OA and solar radiation can better interpret and understand feedbacks between OA and global climatic change." (Page 13 Line 14-15) Deal et al. (2005) reported that the contributions of UVA, UVB and PAR to DMS photodegradation were quite different and Bouillon and Miller (2005) mentioned that pH level can influence the photodegradation of DMS. So the contribution of UVA, UVB and PAR to DMS photodegradation under different pH levels may differ. Furthermore, the ocean undergoes ocean acidification. To better understand the DMS loss through photodegradation in the future ocean, we designed this experiment with five light conditions (natural light, UVA, visible light, dark, UVB: results for full-spectrum natural light minus those for UVB-filtered light) and three pH levels (ambient level: 8.1; level expected at the end of this century: 7.9; level predicted for the middle of the next century: 7.7). "The research of Deal et al. (2005) mentioned that the photodegradation of DMS was wavelength-dependent and the contributions of different wavebands to DMS photodegradation process were quite different. In order to assess the photodegradation process of DMS. . ." (Page 9 Line 17-19)

Question: The authors mention the following on page 3 lines 11-13: "Moreover, we examined the photolysis rate and concentration changes of DMS under the dual stressors of changing pH and solar radiation/temperature and assessed the coupling effects of OA, solar radiation and warming on biogenic dimethylated sulfur compounds cycling." Where is the basic information on light? How was light measured? How did it vary naturally over the 23 day experiment? Where is the basic information on temperature? How was it measured? How did it vary over time? I do not see this anywhere. This is very concerning.

Reply: In this research, photodegradation experiments and temperature experiments were two separate experiments, both of which took 8 hours (Page 4 Line 21-22). The

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light intensity and UV intensity were measured using the agricultural environment detector (Zhejiang Top Instrument Co., Ltd.) and UVR meter (Beijing Normal University Optoelectronics Instrument Factory) and the temperature was detected using thermometer. The variations of light intensity and temperature during experiment were shown in the figure 2. (Page 23 Line 1-4) The relevant content was added in the revised manuscript: “The light intensity and UV intensity were measured using the agricultural environment detector (Zhejiang Top Instrument Co., Ltd.) and UVR meter (Beijing Normal University Optoelectronics Instrument Factory) and the temperature was detected using thermometer. The variations of pH, light intensity and temperature during experiments are shown in Fig. 2.” (Page 4 Line 24-27)

Question: Furthermore, the authors seem to treat light and temperature as a single fixed factor (light/temperature) (? I may be wrong because the text is convoluted and unclear) and proceed with a two-way ANOVA to disentangle the impacts of pH, light, and temperature on concentrations of sulphur compounds? Ecologically and statistically, this is difficult to justify. pH + Light experiments were carried out under natural ambient solar radiation (lines 139-140) with 3 treatments and 1 control (full light) and then information is inferred for a 5th variable by subtracting the effects of one treatment from another treatment. The authors state that they also ran pH + Temperature experiments (with 1 treatment (+4oC) and control but it sounds like these are separate experiments from the light+pH experiments, however it is rather unclear): Lines 140-142: “Temperature experiments were also carried out with unfiltered water in quartz bottle under in situ temperature at 12 oC and high temperature at 18 oC for 8 h. The temperature was continuously controlled by circulating in situ seawater, hot water, and ice.” (This alone begs the question of the uniformity of the temperature treatment itself if ice and hot water are necessary to stabilize the temperature (there were likely a lot of fluctuations, yet these are not reported)). In any case, I have serious doubts as to the validity of lumping together Light and Temperature variables into a single fixed factor as it seems to be proposed in the statistical paragraph of the methodological section. Also are the underlying assumptions of normal distribution for the response variables

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(S compounds) respected? There is no mention of this. The authors state the following at lines 25-28 p.5: “Seawater pH was adjusted with CO₂-saturated seawater during the experiment to maintain a stable pH environment. In the first week, pH level was comparatively stable with a better control, but pH fluctuated obviously and was difficult to control in the stable and decline phases of algal growth. As shown in Fig. 1, the pH showed relatively apparent fluctuations on days 9-12 and days 18-20.” Again, it is difficult here to conceive that pH was a fixed explanatory variable. Judging by Figure 1, there are several instances when there doesn’t appear to be a statistical difference between the pH treatments with very large error bars. On page 5 Lines 14-20, the authors describe very briefly the statistical approaches used, but these spark more questions than answer anything. “Statistical analysis was performed using SPSS version 18.0 (SPSS Inc., IBM, USA). Pearson’s correlation coefficient and probability (p) were calculated to evaluate the quality of the fit when variables were normally distributed. (How was normality assessed?) T-test was used to determine whether a significant difference existed between two treatments. (WHICH treatment? More information is needed.) “Variability in the concentrations of biogenic dimethylated sulfur compounds was analyzed using two-way ANOVA with pH and temperature/light as fixed factors and concentration as a random factor to understand whether an interaction existed between pH and temperature/light on concentration. (Are the authors suggesting that temperature/light are one fixed factor?) Value at $p \leq 0.05$ was considered statistically significant. Linear correlation analyses were used to determine the response of DMS, DMSO concentrations, and bacterial abundance to OA using Origin 9.1.” The authors proceed to establish Pearson’s correlations between two variables within a same pH condition. I am not convinced that this is useful. What is the purpose of this exactly? How does it inform us of the complex response of these variables to fluctuations of pH itself? Overall, the statistical schemes proposed by the authors do not seem appropriate, and they do not facilitate the interpretation of possibly complex responses. Table 4 presents these correlation coefficients. What are the degrees of freedom (DF) for these analyses? Why is a coefficient of 0.791 (between DMSO_d and chl_a) significant at the

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$p < 0.05$ level while a coefficient of 0.778 (between DMSO_d and DMS) is significant at the $p < 0.01$ level? More information is needed about the n and the DF. On pages 10 and 11, the authors discuss these correlations between sulphur compounds and other variables (within a same pH treatment) at length but I am not convinced that these are very informative on the impact of pH itself on the sulphur compounds. When looking at figure 2-3-4-5, the error bars are so wide for the pH treatments (substantial overlapping most of the time, ex: DMSO_p), that it is hard to understand what the purpose of these inner-treatment correlations is. A very convoluted text does not help in the matter. An example of this here: “A significant correlation between DMS and DMSO_d was observed in LC treatments (Table 4). The result was in accordance with the findings of Hatton et al. (2004) and Zindler-Schlundt et al. (2015) who showed that DMSO and DMS are closely related because of the direct formation of DMSO via bacterial DMS oxidation. The close link between DMS and DMSO_d was not observed in MC and HC treatments. The changes in the production, consumption, and degradation processes which caused by the decreasing pH might mask the relationship between DMS and DMSO_d. (Production/consumption and degradation of what exactly?) In addition, DMSP_d concentration showed an obvious correlation with DMSO_d only in LC treatments for the entire duration of the experiment (Table 4). DMSP_d may be cleaved to DMS by specific bacteria that contain DMSP lyase (Curson et al., 2008), followed by the bacterial oxidation of DMS to DMSO_d. This relationship was not found in MC and HC treatments. The phenomenon might be caused by the change in phytoplankton community and bacterial oxidation of DMS to DMSO.” It is very difficult to follow the logic here. What change in phyto community? What are the authors talking about? They do not show any information on community composition for this study. And this is another important shortcoming of this paper: no phytoplankton identification. DMSP/DMS cycling is intimately linked with species composition, yet we have no idea WHO is there and HOW pH may affect the primary producers responsible for a substantial part of the DMS/DMSP cycling. In the introduction the authors mention this: “Shaw hypothesized that DMS and sulfate aerosols are linked to global climate. This link was further

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elaborated by Charlson et al. (1987). Consequently, ocean acidification (OA)-induced changes in the primary productivity might impact on the production rate and sea-to-air emission of DMS and these impacts might further affect cloud formation and climate". The authors do not explore the primary production side of things at all. There are no PP rates, no phyto identification, only chl a which is a proxy of biomass that does not inform us on whether pH impacts chl a levels through physiological processes or through variability in the species composition.

Reply: For the statistical analyses, we used a one-sample Kolmogorov–Smirnov test and Levene test to confirm normal distribution of data and check for homogeneity, respectively. T-test was used to determine whether a significant difference existed between two treatments. Because the conditions during the incubation experiment showed fluctuations, we used the average pH value as the fixed factor and tried our best to estimate the impact of pH on DMS production and release. The analytical method was performed according to the method reported by Arnold et al. (2013). "Arnold, H. E., Kerrison, P., and Steinke, M.: Interacting effects of ocean acidification and warming on growth and DMS-production in the haptophyte coccolithophore *Emiliana huxleyi*, *Global Change Biol.*, 19, 1007–1016, <https://doi.org/10.1111/gcb.12105>, 2013." "degree of freedom" refers to the number of items of data that are free to vary independently. When applied to a set of quantitative data, for a specified value of the mean, only (n-1) items are free to vary. A coefficient of 0.791 (between DMSOd and Chl-a) significant at the $p < 0.05$ level means that DMSOd and Chl-a showed weak correlation in 95% For Table 4, the value of n was 15. We have added this information in the revised manuscript. "Table 4. Pearson's correlation coefficient and associated significance level for mean dimethylated sulfur compounds and Chl-a concentrations under three different CO₂ treatments (n=15)." (Page 34 Line 2) Thanks for the reviewer's comment for sentences "The changes in the production, consumption, and degradation processes which caused by the decreasing pH might mask the relationship between DMS and DMSOd" and "The phenomenon might be caused by the change in phytoplankton community and bacterial oxidation of DMS to DMSO". We agree with

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the reviewer's comment and reworded this sentence in the revised manuscript. "The changes in the conversion of DMSOd to DMS caused by the decreasing pH might mask the relationship between DMS and DMSOd." (Page 11 Line 33- Page 12 Line 1) "This result might be because low pH changed the conversion process among DMSPd, DMS and DMSOd, which broke the relationship between DMSPd and DMSOd. This conclusion further confirmed the inference (section 4.1.1) that OA mainly influenced DMS concentration by affecting the transformation of dimethylated sulfur compounds." (Page 12 Line 4-7) In this research, phytoplankton community was determined only at the beginning and the end of the experiment. At the beginning of the experiment, the phytoplankton community included diatom, dinoflagellate, chrysophyceae, and cryptophyceae with a mean abundance of 2135 cells L⁻¹ (Page 6 Line 13-14). At the end of the experiment, phytoplankton species were less in HC than in LC, and chrysophyceae (e.g., coccolith) and cryptophyceae (e.g., Cryptomonas sp. and Rhodomonas sp.) disappeared during HC treatment (Page 6 Line 23-24). Some previous literature (Zhang et al., 2008) reported that Chl-a was an indicator of phytoplankton biomass, so changes in the phytoplankton biomass were represented by Chl-a concentration during the experiment. "Zhang, H. H., Yang, G. P., Zhu, T.: Distribution and cycling of dimethylsulfide (DMS) and dimethylsulfoniopropionate (DMSP) in the sea-surface microlayer of the Yellow Sea, China, in spring, *Contin. Shelf Res.*, 28, 2417–2427. <https://doi.org/10.1016/j.csr.2008.06.003>, 2008"

Question: 1.4 Out-dated information and statements that are too general. Page 1 line 28. The information given here is not up to date. The authors reference a paper written in 2000. Progress has been made in the last ca. 15 years. Anthropogenic carbon dioxide (CO₂) emissions have increased atmospheric CO₂ concentrations from their pre-industrial value of 280 to ca. 400 μatm in 2016 (NOAA-ESRL). Page 1 lines 29-30. Under which scenario exactly? Please be precise and use the latest information available. Concentrations of 850 to 1370 μatm are expected by the end of the century under the business-as-usual scenario RCP 8.5 (IPCC, 2013). Page 1 line 30 and beyond. The authors don't offer any explanation, even if short, as to the underlying

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processes involved in pH modulation in oceans as a result of CO₂ increases in the atmosphere. They should spend a little more time explaining this. That is one the focuses of their paper after all. Also there is a more recent paper by Caldeira and Wickett (2005) suggesting that the surface ocean pH is expected to decrease by an additional 0.3-0.4 units under the RCP 8.5 scenario by 2100. Page 2 lines 1-2. What is the reference here? There are several consequences of increasing OA. Modifications in DIC is the primary, from which modifications in carbonate system ensue (which includes modifications in saturation states of calcite and aragonite, which by the way the authors don't explain the importance of: why talk about calcite and aragonite?). Modifications in the amount of protons (H⁺) is another. Etc. Etc. Page 2 lines 2-4. This is quite a general statement. What changes exactly (less calcite, more H⁺, more CO₂?) will affect what physiological processes exactly, and what type of marine "organism" exactly? There is so much literature on the many aspects of OA and the consequences for marine organisms (Calcifiers? Phyto? Zoo? Fish? Etc), and the potential impacts may be positive, negative, resilience etc. . . The phrase written is simply too generalist, it should offer at least a glimpse into the multi-tiered discoveries made by the OA research community, or at least focus on the aspects that are relevant to the author's research: DMS-producing microbial communities. . . Page 2 lines 19-20. The authors make very general statements without offering much information: "The conversion of DMSP to DMS is controlled by a number of chemical and biological processes." Yes. But which ones? These are only some examples of the generalities, more can be found in the manuscript.

Reply: Thanks for the reviewer's comment. We have rephrased the relevant content and updated new literature in the manuscript. "The current atmospheric CO₂ concentration (pCO₂) is around 400 μatm (NOAA-ESRL) and rising at an unprecedented rate. Concentrations of 850 to 1370 μatm are expected by the end of the century under the business-as-usual scenario RCP 8.5 (IPCC, 2013). The oceanic uptake of anthropogenic CO₂ emissions is leading to a change of seawater carbonate system, manifested as increasing protons [H⁺], falling [CO₃²⁻] and a drop in seawater pH. Equi-

libration of seawater with increasing CO₂ concentration in the atmosphere has already declined the surface ocean pH levels by 0.3–0.4 units compared to pre-industrial values (Caldeira and Wickett, 2005). The primary consequence of the inexorable increase in oceanic acidity is a change in seawater carbonate system. This change could affect marine ecological environment and primary productivity, as low pH will affect physiological process of marine phytoplankton (Fu et al., 2007; Mélançon et al., 2016; Orr et al., 2005). For example, pH conditions can influence the intracellular acid-base balance and the energy demand of phytoplankton (Kramer et al., 2003). Physiological activities of phytoplankton may be influenced, and therefore may affect the production and release of biogenic compounds, such as marine biogenic trace gas. The oceans are an important source of some atmosphere trace gases which affect atmospheric chemistry and global climate (Arnold et al., 2013; Asher et al., 2016; Park et al., 2014). Change in surface ocean pH will have an important impact on the climate.” (Page 1 Line 28-30, Page 2 Line 1-10) “Caldeira, K., and Wickett, M. E.: Oceanography: anthropogenic carbon and ocean pH, *Nature*, 425, 365–365, <https://doi.org/10.1038/425365a>, 2003” has replaced with “Caldeira, K., and Wickett, M. E.: Ocean model predictions of chemistry changes from carbon dioxide emissions to the atmosphere and ocean, *J. Geophys. Res. Oceans*, 110, <https://doi.org/10.1029/2004JC002671>, 2005.” (Page 15 Line 30-31) “The conversion of DMSP to DMS is controlled by a number of chemical and biological factors (Archer et al., 2002), such as activity of DMSP-lyase enzymes (Stefels et al., 2000) and condition of light (Slezak et al., 2007).” (Page 2 Line 22-24) “Slezak, D., Kiene, R. P., Toole, D. A., Simó, R., Kieber, D. J.: Effects of solar radiation on the fate of dissolved DMSP and conversion to DMS in seawater, *Aquat. Sci.*, 69, 377–393, <https://doi.org/10.1007/s00027-007-0896-z>, 2007.” (Page 19 Line 28-29) “Stefels, J.: Physiological aspects of the production and conversion of DMSP in marine algae and higher plants, *J. Sea Res.* 43, 183–197, [https://doi.org/10.1016/S1385-1101\(00\)00030-7](https://doi.org/10.1016/S1385-1101(00)00030-7), 2000.” (Page 20 Line 7-8)

Question: 2. On the form As it currently stands, the overall level of language does not meet the high standards of Limnology and Oceanography There are countless

examples of awkward formulations and missing words throughout the paper, including: “. . .more CO₂ (low pH) will affect physiological process of marine organism.” Page 2 line 3 Page 2 lines 5 and beyond: The transition between the impact of OA on they physiology of marine organisms and the oceanic production of climate-relevant gases is not well established.

Reply: In order to meet the high standards of Biogeosciences, we have modified the whole article and the sentence “. . .more CO₂ (low pH) will affect physiological process of marine organism” has been replaced with “. . .low pH will affect physiological process of marine phytoplankton. . .” (Page 2 Line 5) “This change could affect marine ecological environment and primary productivity, as low pH will affect physiological process of marine phytoplankton (Fu et al., 2007; Mélançon et al., 2016; Orr et al., 2005). For example, pH conditions can influence the intracellular acid-base balance and the energy demand of phytoplankton (Kramer et al., 2003). Physiological activities of phytoplankton may be influenced, and therefore may influence the production and release of biogenic compounds, such as marine biogenic trace gas. The oceans are an important source of some atmosphere trace gases which affect atmospheric chemistry and global climate (Arnold et al., 2013; Asher et al., 2016; Park et al., 2014). Change in surface ocean pH will have an important impact on the climate.” (Page 2 Line 4-10)

Question: Once emitted to atmosphere, DMS. . .”. Page 2 line 10 “DMSO was initially conceived as a sink for DMS. Nevertheless, DMSO was later found a potential source of DMS.” Page 2 lines 26-27 “The ocean undergoes multiple environmental changes. Other climatic ecological stressor or factors would probably alter the effects of ocean acidification on the production and consumption process of DMS in both direct and indirect ways.” Page 3 lines 2-4 “The photochemical process of DMS in the surface water would change due to the changing light level and seawater pH level.” Page 3 line 5 There are so many more examples. I will stop here because I believe a profound language editing needs to be conducted and this goes beyond the scientific mandate of a reviewer. The introduction offers many general statements that only skim a frac-

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tion of the inherent complexity and wealth of information related to OA research. The paper lacks coherence and clarity and there is redundancy in the writing. Formulation of phrases is awkward, making the text very difficult to follow. Page 2 lines 6-8: Redundancy in the phrase: “It is produced by enzymatic cleavage of dimethylsulfoniopropionate (DMSP) (Gabric et al., 2010), which is synthesized by marine phytoplankton as a phytoplankton-derived precursor of DMS.” Page 2 lines 16-17: Redundancy in the phrase as well as unclear: “DMSP, as the main precursor of DMS, . . ., presents an important effect on the biogeochemical cycle of climatically active trace gas DMS.” This is rather vague and redundant. Needs rephrasing.

Reply: According to the reviewer’s suggestion, we have revised the relevant sentences in the revised manuscript. The sentence “It is produced by enzymatic cleavage of dimethylsulfoniopropionate (DMSP) (Gabric et al., 2010), which is synthesized by marine phytoplankton as a phytoplankton-derived precursor of DMS.” and the reference “Gabric, A. J., Cropp, R., Hirst, T., and Marchant, H.: The sensitivity of dimethyl sulfide production to simulated climate change in the Eastern Antarctic Southern Ocean, *Tellus*, 55, 966–981, <https://doi.org/10.1034/j.1600-0889.2003.00077.x>, 2010” have been deleted. The sentence “Nevertheless, DMSO was later found a potential source of DMS (Hatton et al., 2012). DMSO can be biologically reduced to DMS via enzymatic reactions that might depend on reductases (Spiese et al., 2009). Thus, DMSO is a key compound in the complex redox loop that is involved in marine sulfur cycle because it can be both an end product of DMS oxidation and a precursor of DMS, which potentially plays an important role in climate regulation (Charlson et al., 1987; Deschaseaux et al., 2014)” has been replaced with “DMSO and DMS can be converted to each other through photochemical and biological oxidation processes (Brimblecombe and Shooter, 1986; Toole and Siegel, 2004; Hatton et al., 2012; Spiese et al., 2009). Thus, DMSO is a key compound in the complex redox loop that is involved in marine sulfur cycle, which potentially plays an important role in climate regulation (Charlson et al., 1987; Deschaseaux et al., 2014).” (Page 2 Line 30-33)

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Question: Page 2 line 21 The word “preferred” is not appropriate.

Reply: We agree with the reviewer’s comment and have deleted the “preferred and” in the manuscript.

Question: Page 2 line 26: “DMSO was originally conceived as a sink for DMS.” This phrase does not make sense. Needs rewording. Again, these are only examples, more can be found in the manuscript. In short, at this point, there is much work to be done before this paper can be considered for publication in Limnology and Oceanography. First and foremost, there are several uncertainties and questions related to the methodologies used and the experimental design itself. Without a sound experimental plan and measurements, the rest of it (interpretations and conclusions) is useless. Overall, the entire methodological aspects related to the OA part of the experiments are unclear and lacking (carbonate system measurements absent, salinity?, temperature records?). It is unclear whether the temperature and light treatments were separate experiments. And where are those measurements of temperature and light? What about primary production, phytoplankton species composition? The statistical schemes do not seem appropriate. The objectives of the study are not well stated (I am referring here to the Estuarine context in which the experiment takes place). The environmental setting is not clearly established; the authors don’t even situate the study area on a map. The level of language is not up to par.

Reply: According to the reviewer’s suggestion, we have changed “DMSO was originally conceived as a sink for DMS” to “DMSO can also be directly synthesized by phytoplankton and plays similar biological functions to DMSP (Gao et al., 2017). DMSO and DMS can be converted to each other through photochemical and biological oxidation processes (Brimblecombe and Shooter, 1986; Toole and Siegel, 2004; Hutton et al., 2012; Spiess et al., 2009)” in the revised manuscript. (Page 2 Line 29-31) “Gao, N., Yang, G. P., Zhang, H. H., Liu, L.: Temporal and spatial variations of three dimethylated sulfur compounds in the Changjiang Estuary and its adjacent area during summer and winter, Environ. Chem., 14, 160–177, <http://dx.doi.org/10.1071/EN16158>,

2017.” (Page 17 Line 1-3) Yes, the temperature and light treatments were separate experiments. In order to better understand, we explained it in the revised manuscript and added the graph of temperature and light intensity in Figure 2. “In the separate photodegradation experiment,..” (Page 4 Line 12) “Another separate experiment of temperature was also carried out with unfiltered water in quartz bottle under in situ temperature at 12 °C and high temperature at 18 °C for 8 h.” (Page 4 Line 21-22) Unfortunately, we measured the initial temperature (12.33 °C), salinity (33.79‰, and carbonate parameters of three treatments only at the beginning of the experiment, the parameters during the experiments were not determined. Thanks for the reviewer’s comments and we will monitor these parameters in real time in our future research. In order to make the study area more clearly, we have marked the sampling station on a map (see Figure 1). (Page 22 Line 1-2)

Please also note the supplement to this comment:

<https://www.biogeosciences-discuss.net/bg-2017-453/bg-2017-453-AC3-supplement.pdf>

Interactive comment on Biogeosciences Discuss., <https://doi.org/10.5194/bg-2017-453>, 2017.

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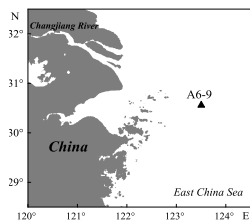


Figure 1. Location of the sampling station (123.50°E, 30.56°N) during the cruise.

Fig. 1. Figure 1

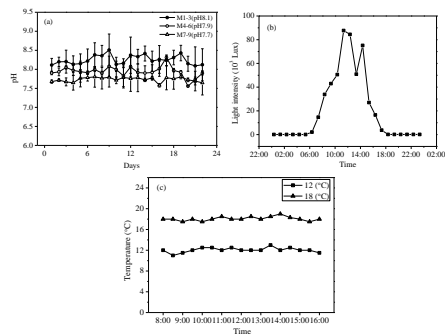


Figure 2. Temporal changes in pH (a) for averages of the nine barrels over the triplicates treatments during ocean acidification incubation experiment, variations of light intensity (b) during photodegradation experiment and temperature (c) during temperature experiment.

Fig. 2. Figure 2

Table 1. Preliminary carbonate parameters of three treatments during the incubation experiment.

Treatments	pH \pm s.d.	DIC \pm s.d. ($\mu\text{mol kg}^{-1}$)	pCO ₂ \pm s.d. (μatm)	HCO ₃ ⁻ \pm s.d. ($\mu\text{mol kg}^{-1}$)	CO ₃ ²⁻ \pm s.d. ($\mu\text{mol kg}^{-1}$)
M1-3	8.1 \pm 0.008	2271 \pm 0.577	389.2 \pm 5.661	2087.2 \pm 3.371	168.0 \pm 3.926
M4-6	7.9 \pm 0.012	2263 \pm 1.000	635.9 \pm 13.80	2130.0 \pm 4.133	107.1 \pm 4.093
M7-9	7.7 \pm 0.032	2274 \pm 1.154	1040.7 \pm 54.05	2164.0 \pm 4.193	67.5 \pm 7.552

Fig. 3. Table 1

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